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(54) Title: BETA-AMYLOID TOXICITY

(57) Abstract

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The present invention relates to methods and compositions for visualization of the toxic effects of transgenes in vivo. In particular, the present invention provides methods and compositions for the production and use of transgenic, including dually transgenic, Caenorhabditis elegans for visualization of the toxic effects of β -amyloid accumulation in vivo.

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BETA-AMYLOID TOXICITY

FIELD OF THE INVENTION

The present invention is in the field of medicine and molecular biology. In particular, the invention provides transgenic non-human animals in which expression of a reporter gene is induced by a toxic transgene. The invention also provides methods and compositions for *in vivo* visualization of the toxicity associated with toxic proteins. In addition, the present invention provides methods and compositions for the production and use of transgenic non-human animals for screening candidate drugs to assess and treat toxicity.

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BACKGROUND OF THE INVENTION

Investigation of the molecular mechanisms of various diseases has suggested that many pathologies are caused by expression of proteins that are immediately or gradually deleterious to the cells where they are expressed. Among these diseases are Alzheimer's disease, prion diseases, Huntington's disease, and amyotrophic lateral sclerosis. Several of these diseases are thought to result from aberrant folding of proteins, which results in the accumulation of toxic proteins or protein aggregates.

Alzheimer's disease is associated with the majority of dementia cases in the United States, with an estimated 2 million people afflicted with the disease, and a mortality rate of approximately 100,000 people per year (See, R.W.P. Cutler, "Degenerative and Hereditary Diseases." in Medicine, Scientific American, New York, (1988), pages 11 (IV):1-13; and R. Katzman (1986) N. Engl. J. Med. 314:964). It has been estimated that the total cost for nursing home care alone of Alzheimer's patients exceeds \$13 billion/year (See, M.M. Heckler (1985) Am. Psychol. 40:1240). According to the Centers for Disease Control (CDC), mortality due to Alzheimer's disease in the United States increased 10-fold between 1979 and 1987 (0.4 per 100,000 to 4.2 per 100,000) (See, "Reported death rate for Alzheimer's is up tenfold since 1979," (1990) Clin. Psychiatr. News 18:21).

Patients suffering from Alzheimer's disease typically suffer progressive memory deficit, progressive decline in cognitive functions, anxiety, depression, visuospatial and speech deficits, delusions, personality changes, motor skill deterioration, loss of verbal

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ability, and incontinence. Eventually, patients are completely incapacitated and disoriented, requiring total care. The course of Alzheimer's disease ranges from less than three years to over 20 years before death occurs. However, in typical cases, it progresses at a fairly constant rate, with an average duration of 6 to 10 years.

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Alzheimer's disease is not the only cause of dementia. Indeed, there are over fifty recognized causes of dementia. As some causes of dementia are amenable to treatment, differential diagnosis of patients suffering dementia is particularly important. Because at least 20% of clinically diagnosed patients were found at autopsy to have had conditions other than Alzheimer's disease, the National Institute of Neurological and Communicative Disorders and Stroke (NINCDS) and Alzheimer's Disease and Related Disorders Association (ADRDA) retined the clinical diagnostic criteria for Alzheimer's disease (*See.* M.A. Jenike. "Psychiatry." *in Medicine*, Scientific American, New York, [1991], pages 13 (V):1-5). Based on these criteria, the diagnosis of Alzheimer's disease may be "definite" (*i.e.*, requiring examination of brain tissue), "probable" (*i.e.*, patients have deficits in two or more areas of cognition, insidious onset of disease, progressive worsening of memory and other cognitive functions, and normal consciousness levels), or "possible" (*i.e.*, patients meet the criteria for probable Alzheimer's disease, but exhibit variations in the disease course or have a systemic illness that is sufficient to cause dementia, but is not considered to be the cause of the dementia).

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The numerous varieties of dementia, and variations in patient presentations, often make diagnosis problematic. Thus, the NINCDS/ADRDA criteria are very detailed, and necessitate the thorough examination of patients with suspected Alzheimer's disease. Currently, the only way to obtain a definite diagnosis is by post-mortem histological examination of brain tissue for the presence of senile plaques.

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The histopathological lesions of Alzheimer's disease include neuritic or senile plaques, neurofibrillary degeneration, and granulovacuolar neuronal degeneration. The senile plaques usually contain a core of insoluble, amyloidic extracellular material (" β -amyloid) surrounded by a halo of neurofibrillary tangles and dystrophic neurons. The primary protein component of the amyloidic core of senile plaques is a 4.2 kd amyloid β peptide (often referred to as " $A\beta$ "). A number of other proteins have also been identified

as components of senile plaques, including anti-chymotrypsin and apolipoprotein E. The major component of the neurofibrillary tangles is an abnormally phosphorylated microtubule-associated protein referred to as "tau." The β-amyloid protein in senile plaques is a small glycoprotein (*i.e.*. a 39-43 amino acid protein, derived from the larger amyloid precursor protein), that has been detected in non-neural tissues (*e.g.*, skin, subcutaneous tissue, and intestines) and blood vessels of Alzheimer's disease patients (*See*, Cutler, *supra*). Deposits of amyloid may be detected by their ability to bind specific dyes, such as Congo red or thioflavin S. Thus, it has been further hypothesized that detection of this protein may serve as a potential diagnostic aid in the assessment of patients suffering from dementia.

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Alzheimer's disease usually arises spontaneously, although genetics play a role in development of the disease. For example, the histopathologic lesions of Alzheimer's disease have been regularly observed in the brains of older patients dying of Down's syndrome (*See*, Cutler, *supra*). It is known that the β-amyloid gene resides on chromosome 21. As Down's patients have an extra copy of this chromosome, there is an increased expression of this chromosome in these patients. It has been hypothesized that increased expression of the β-amyloid gene may regulate the formation of amyloid plaques ("senile plaques") in these patients. In addition, four loci were recently identified as playing a role in the genetic susceptibility of Alzheimer's disease (*See*, Pericak-Vance and Haines (1995) *Trends Genet.* 11:504).

Other potential risk factors for the development of Alzheimer's disease include environmental factors (e.g., head trauma, smoking, and exposure to heavy metals), sociological factors (e.g., depression and educational level), biological factors (e.g., increasing age and hyperthyroidism), and a family history of Alzheimer's disease, Down's syndrome, or Parkinson's disease (See. Pericak-Vance and Haines, supra). Nonetheless, despite recent advances, the exact etiology and pathogenesis of Alzheimer's disease remain largely unknown.

In addition to the problems associated with diagnosing Alzheimer's disease. improvements are needed in the area of treatment. Many agents have been tested for their ability to treat the cognitive decline associated with Alzheimer's disease. For example,

various cholinergic enhancers (e.g., choline and lecithin) have been tested. Unfortunately, cholinergic precursors have been shown to be not useful, although some drugs that stimulate cholinergic transmission may be helpful in some patients. One example is physostigmine, a compound that prevents the synaptic breakdown of acetylcholine. However, the overall clinical effect of this drug has not been as dramatic as initially hoped (See, M.A. Jenike, "Psychiatry," in Medicine, Scientific American, New York, (1991), pages 13 (V):1-5).

Other drugs, such as tetrahydroaminoacridine (THA or tacrine), a centrally acting anti-cholinesterase, have been tested. In a large multi-center trial of THA, liver enzyme abnormalities were reported and the preliminary results indicated that, at least at low dosages. THA is not an effective treatment of Alzheimer's disease (Jenike, *supra*). Ergoloid mesylates (Hydergine) is an extremely safe compound, and remains the most commonly prescribed drug for patients with Alzheimer's disease. However, the overall effects of the drug are at best minimal.

The mechanisms of toxicity due to accumulation of β-peptide are currently the subject of much investigation, and no definitive causes of toxicity have yet been established. (See, Benzi and Moretti (1995) Neurobiology of Aging, 16:661-674). Because of this uncertainty, efforts to develop mechanism-based treatment regimens have not been possible. Compounds presently in use to treat Alzheimer's disease only serve to alleviate the systemic effects associated with the disease.

In order to study disease mechanisms and genetic-based phenomena, animals in which a foreign gene has been inserted have been described by various researchers. International Patent Application WO 96/03034 describes insertion of retroviral vectors into fish, in order to produce fish with desirable traits or to study development. Various transgenic animal models for Alzheimer's disease are described in International Patent Applications WO 93/14200, WO 93/02189, WO 94/12627, WO 94/23049, and European Patent Publication EP 653154. Typically these transgenic animals are mice or other mammals; however, β-amyloid peptide has been expressed in the nematode *Caenorhabditis elegans* (Link (1995) *Proc. Natl. Acad. Sci.* 92: 9368). The use of *C. elegans* for investigation of mutant forms of the *C. elegans* genes mec-4 and deg-1, which

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cause neurodegeneration, is described in U.S. Patent No. 5,196,333. Drawbacks to mammalian animals as model systems are the relatively long generation time, which makes mammals less desirable for high-throughput screening of potential pharmaceuticals, and the difficulty in studying the molecular processes of interest without sacrificing the animal for cell and tissue analysis.

Reporter genes are genes that encode proteins or other compounds that can be detected by a variety of methods, and which "report" the occurrence of successful introduction and expression of gene sequences. β-galactosidase and luciferase are examples of such reporter genes. Recently, the gene for green fluorescent protein (GFP) of the jellyfish *Aequorea victoria* has been described in U.S. Patent No. 5,491,084. This gene can provide a method for indicating expression of a gene via fluorescent detection of GFP. The drawback to using GFP in mammalian systems is as described above, that is, the need to sacrifice the mammal in order to analyze the cells and tissues of interest. This drawback is especially severe in the context of large-scale screening of potential therapeutic compounds, and U.S. Patent No. 5,491,084 does not describe a system suitable for high-throughput screening of pharmaceuticals for activity against toxic proteins expressed in cells.

Thus, methods and animal systems are needed to screen drugs quickly and inexpensively for their effects on proteins and other substances associated with Alzheimer's disease. Convenient methods and animal systems for screening drugs for other neurodegenerative diseases, such as prion diseases, Huntington's disease, and amyotrophic lateral sclerosis, are also desirable.

SUMMARY OF THE INVENTION

The present invention provides methods and compositions useful for the production and use of dually transgenic animals, in particular *Caenorhabditis elegans*. In addition, in one embodiment, the present invention provides transparent animals that express a reporter gene inducible by a toxic transgene. In a preferred embodiment, the toxic transgene encodes β -peptide. Although it is not intended that the present invention be limited to any particular reporter, in a preferred embodiment, the reporter gene is green fluorescence

protein (GFP). It is contemplated that the effects or product of the reporter gene be observable (*i.e.*, the reporter gene is expressed). For example, in embodiments in which GFP is the reporter, its presence is detected using fluorescence microscopy. In addition, in these embodiments, the animals do not need to be sacrificed in order to observe the expression of the reporter gene (*i.e.*, the detection may be accomplished on living animals). If other reporters are used, other detection methods may be necessary. For example, lacZ expression may be detected by exposing the tissues of the animal to the substrate for the gene (*i.e.*, β -galactoside), and observing for the presence of blue dye in the tissues. However, this method requires that the animal be sacrificed in order to observe the expression of the reporter gene.

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In one alternate embodiment, the genome of the transparent animal further contains a heterologous marker gene. In a preferred embodiment, the marker gene is rol-6(su-1006). In the embodiments in which rol-6(su-1006), expression of the marker gene is accomplished by observing the living animals for the rolling behavior that results due to the expression of the abnormal collagen gene. In contrast, wild-type animals exhibit normal locomotion and do not exhibit this rolling behavior, as their collagen is normal.

In an alternative embodiment, the first transgene of the dually transgenic non-human animal comprises pCL25. In another alternative embodiment, the second transgene of the dually transgenic non-human animal comprises pCL12. In one alternate embodiment, the genome of the transparent animal further contains a heterologous marker gene. In a preferred embodiment, the marker gene is rol-6(su-1006). In the embodiments in which rol-6(su-1006), expression of the marker gene is accomplished by observing the living animals for the rolling behavior that results due to the expression of the abnormal collagen gene. In contrast, wild-type animals exhibit normal locomotion and do not exhibit this rolling behavior, as their collagen is normal.

In a preferred embodiment, the transparent animal is selected from the class Nematoda. Although it is not intended that the transparent animal of the present invention be limited to any specific animal, in a particularly preferred embodiment, the transparent animal is *Caenorhabditis elegans*.

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In an alternate embodiment the cells of the transparent animal display toxicity resulting from the accumulation of β -peptide within the cells. In one preferred embodiment, the genome of the transparent animal comprises SEQ ID NO:5 and SEQ ID NO:8.

The present invention also provides methods for producing dually transgenic non-human animal comprising: providing: a first and second non-human animal; a first transgene comprising β -peptide: and a second transgene, comprising a reporter: introducing the first transgene into the genome of the first non-human animal to produce a first transgenic animal, and introducing the second transgene into the genome of the second non-human animal to produce a second transgenic animal; and mating the first transgenic animal with the second transgenic animal to produce a dually transgenic animal, wherein the β -peptide and the reporter are expressed.

In one embodiment of the methods of the present invention the dually transgenic non-human animal is transparent. In a preferred embodiment, the transparent animal is a nematode, while in a particularly preferred embodiment, the animal is *Caenorhabditis* elegans.

In an alternative embodiment, the first transgene of the dually transgenic non-human animal comprises pCL25. In another alternative embodiment, the second transgene of the dually transgenic non-human animal comprises pCL12. In one alternate embodiment, the genome of the transparent animal further contains a heterologous marker gene. In a preferred embodiment, the marker gene is rol-6(su-1006). In the embodiments in which rol-6(su-1006), expression of the marker gene is accomplished by observing the living animals for the rolling behavior that results due to the expression of the abnormal collagen gene. In contrast, wild-type animals exhibit normal locomotion and do not exhibit this rolling behavior, as their collagen is normal.

The present invention also comprises methods for testing compounds for anti-toxic effects, comprising: providing a dually transgenic non-human animal expressing a toxic transgene and a reporter; a composition comprising a test compound in a form suitable for administration such that the compound is bioavailable in the cells of the animal; and administering the test compound to the non-human animal. The toxic transgene induces

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expression of the reporter gene. In a preferred embodiment, the toxic transgene is β -peptide and the reporter gene is GFP. In one embodiment, the method further comprises the step of measuring a reduction or increase in the expression of the reporter by the dually transgenic non-human animal and thereby identifying a compound as therapeutic. In a particularly preferred embodiment of the methods, the compounds inactivate the β -peptide expressed by the dually transgenic animal.

In one embodiment of the methods for testing compounds for β-peptide toxicity, the dually transgenic non-human animal is transparent. In a preferred embodiment, the transparent animal is a nematode, while in a particularly preferred embodiment, the animal is *Caenorhabditis elegans*. In an alternative embodiment, the first transgene of the dually transgenic non-human animal comprises pCL25. In another alternative embodiment, the second transgene of the dually transgenic non-human animal comprises pCL12. In one alternate embodiment, the genome of the transparent animal further contains a heterologous marker gene. In a preferred embodiment, the marker gene is *rol-6(su-1006)*, which produces effects described herein.

In one alternative embodiment, one or more test compounds are tested for their ability to counter the toxic effects of transgene product. It is also contemplated that the test compounds will be tested for their ability to prevent the expression of the toxic transgene, for example, β -peptide.

In alternative embodiments, dually transgenic animals in which the expression of, or effects of transgene toxicity, are reduced or eliminated by the test compounds, are mated. The progeny of these matings are also then tested for the effects of test compounds on the expression of, or effects of toxicity. In yet another embodiment, the progeny of these matings are used in other assay systems for the identification of therapeutic compounds.

In yet another alternative embodiment, the dually transgenic animals of the present invention are used to identify methods suitable for the diagnostic testing of Alzheimer's disease or pathology due to other diseases. Thus, the dually transgenic animals of the present invention are used to develop assays suitable for use in humans or animal models of Alzheimer's disease.

DESCRIPTION OF THE FIGURES

Figure 1 is a diagram illustrating the construction of dual transgenic animals expressing β -amyloid peptide and an hsp/GFP reporter.

Figure 2 shows the DNA sequence (SEQ ID NO:5) and restriction map for pCL12. Figure 3 is a graphic map of pCL12.

Figure 4 shows the DNA (SEQ ID NO:6), and amino acid sequence (SEQ ID NO:7) of pCL12 from nucleotide 1071 through 1253 (i.e., the β -(1-42) nucleic acid and amino acid sequence).

Figure 5 shows the DNA sequence (SEQ ID NO:8) and restriction map for pGFP-TT.

Figure 6 is a graphic map of pGFP-TT.

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Figure 7 shows the DNA sequence (SEQ ID NO:9) and restriction map for pCL25.

Figure 8 is a graphic map of pCL25.

Figure 9 shows the DNA sequence (SEQ ID NO:10) of rol-6.

Figure 10 shows the amino acid sequence (SEQ ID NO:11) of rol-6.

DESCRIPTION OF THE INVENTION

The present invention provides methods and compositions useful for the production and use of transgenic animals. The methods and transgenic animals of the invention also provide an efficient and effective system for screening drug effective in ameloriating the effects of toxic gene products.

The practice of the present invention will employ, unless otherwise indicated, conventional techniques of molecular biology, microbiology, cell biology and recombinant DNA, which are within the skill of the art. See, e.g., Sambrook, Fritsch, and Maniatis, MOLECULAR CLONING: A LABORATORY MANUAL, 2nd edition (1989); CURRENT PROTOCOLS IN MOLECULAR BIOLOGY, (F.M. Ausubel et al. eds., 1987); the series METHODS IN ENZYMOLOGY (Academic Press, Inc.); PCR 2: A PRACTICAL APPROACH (M.J. McPherson, B.D. Hames and G.R. Taylor eds., 1995); ANIMAL CELL CULTURE (R.I.

Freshney, Ed., 1987); and ANTIBODIES: A LABORATORY MANUAL (Harlow et al. eds., 1987).

Animals

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The methods and systems described herein can be practiced with any non-human animal. In a preferred embodiment, the animal used is transparent for at least part of its life cycle, for example, oocytes (*Xenopus* or others); larvae; pupae, fish (zebrafish or others); and nematodes, such as *C. elegans*. *C. elegans* is particularly preferred because it is transparent throughout its life and has been extensively studied as a model to determine developmentally regulated gene expression, as well as pattern formation. The visibility of all of the animals' cells throughout their life cycles, has led to the complete determination of the cell lineages, and a detailed description of the morphogenic changes that occur during *C. elegans* development. Rapid methods for introduction of cloned DNA into the *C. elegans* germ line has provided means to study molecular function and expression *in vivo*, as the effects of the gene expression are usually readily visible in the transparent worms.

Reporter Molecules

The methods and compositions of the present invention make use of reporter genes to monitor gene activity. Visible and quantifiable reporter genes are known and described in the art. Successfully used reporter molecules in gene fusion vectors in studies with *C. elegans* include the *E. coli lacZ* coding region (*See e.g.*, A. Fire *et al.*, (1990) *Gene* 93:189-198), and the *Aequora victoria gfp* coding region (*See e.g.*, M. Chalfie *et al.*, (1994) *Science* 263:802-805; D.C. Prasher *et al.*, (1992) *Gene* 111:229-233; and Genbank Accession #M62654), which produces an intrinsically fluorescent protein. Although various fusion expression vectors have been used and reported in the literature, problems have been encountered (*See e.g.*, A. Fire, "Fire Lab Vector Kit"--June 1995). For example, with *lacZ*, there have been many reports in which it was not possible to correlate transgene expression patterns with physiological expectations, or only weak correlations have been possible. Ectopic expression is often frequently seen with short promoters, and occurs

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most prominently in the gut and pharynx of the affected animals. This may be due to weak promoter and/or enhancer signals in the vectors. Expression pattern deficits of *lacZ* fusions have been classified into three groups. In the first, transgenes are expressed in the correct tissue, but mosaic expression is observed (*i.e.*, only a subset of the cells stain during the detection methods). This has been observed even with integrated high-copy transgenes. In the second group, expression in a single tissue or cell population is not seen with the transgene. Finally, there has been the failure of transgene fusion constructs to show expression in the pre-12 cell embryo, or in any embryonic, larval, or adult germ line.

The gfp fusion vectors were developed as an alternative to the lacZ markers, but have been available for a much shorter time (See e.g., Chalfie et al., (1994), supra; U.S. Patent No. 5.491,084). Initial reports indicate that the fluorescence pattern appears to be more restricted than that exhibited by equivalent lacZ fusion. In addition, there are still problems associated with the germline expression of the transgenes and ectopic expression. Indeed, in some cases, the problems with ectopic expression have been exacerbated by these "improvements."

The present invention overcomes many of these problems described in the art. The transparent animals and dually transgenic animals described herein provide easily visible, stably expressed systems in which the toxic transgene and reporter gene are expressed similarly to endogenous chromosomal gene expression. These animals provide an excellent system for screening compounds having effects on the toxic transgene. The present invention, therefore, provides model systems for the study of human diseases and methods of identifying therapeutic compounds using these animal systems.

Inducible Promoters

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As discussed above, the reporter genes are operably linked to an inducible promoter. The promoter is induced by the toxic transgene, for example by the gene product of the toxic transgene. It is also contemplated that the toxic transgene can act to induce the promoter indirectly, for example by disrupting other cellular proteins or functions. Suitable inducible promoters are available and can be readily determined by those skilled in the art. Non-limiting examples of promoters which are induced by

"stresses" include the metallothionein gene promoter (*mtl*-1 or *mtl*-2; *e.g.*, Genbank Accession #M92910, #M11794, #X00504, and #X00953), and the *C. elegans* amyloid precursor protein (APP) homolog *apl*-1 gene promoter (*See e.g.*, Daigle and Li, Proc. Natl. Acad., Sci., 90:12045-12049 [1993]). Other potential promoters include those from other inducible heat-shock genes; at least one of the known *C. elegans hsp*70 genes is strongly heat-inducible. It is also contemplated that promoters from genes known to be upregulated under stress conditions in other systems (*e.g.*, superoxide dismutase, catalase, glutathione reductase, etc.) may also be useful.

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In one embodiment, as described in the Examples below, a strong muscle-specific promoter was used to express a potentially secretable form of the β -peptide, so as to generate significant extracellular levels of β -peptide. This was accomplished in order to mimic the situation that may exist in the human brain, and allow observations of cell-external neurotoxicity. The present inventor has also demonstrated that the promoter must be chosen so that enough β -peptide is produced to cause physiological effects, but not so much to kill the animals.

The unc-54/ β -(1-42) minigene was constructed with a modified signal sequence that has been previously shown to allow secretion of a her-1 protein product that is ectopically expressed in muscle cells (M.D. Perry et al., (1993) Genes Dev., 7:216-228). The development of C elegans transgenic for expression of β -amyloid was previously described by the inventor (Link, (1995) Proc. Natl. Acad. Sci., 92:9368-9572).

However, in early experiments, it was observed that β -peptide deposits were not convincingly detected outside of the muscle cells, when tested with the antibodies described in Example 4, below. While an understanding of the mechanism is not necessary for the practice of the present invention, it is apparent that the majority of β -peptide expressed by the transgenic worms is retained in the muscle cells and is responsible for the pathology observed in the muscle cells.

It is also contemplated that β -peptide expression will be directed to other tissues, through utilization of appropriate promoters. For example, it is contemplated that animals expressing β -peptide in the intestine may be particularly useful to analyze compounds such

as drugs for their effect on β -peptide, as these cells readily take up exogenous compounds administered orally.

Transgenes

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The transgene element of the present invention can be any sequence which is able to induce the promoter operably linked to the reporter gene. As noted, the transgene is "toxic" in the sense that it disrupts cellular function in some way. Preferably, the transgene encodes a protein that is toxic to the host cells and/or organism in that it causes deleterious effects to the host, for instance interfering with the hosts ability to survive and/or grow. The toxic transgenes encode proteins that disrupt cellular function directly (e.g., the gene product is toxic) or indirectly (e.g., the sequence of the transgene disrupts cellular function by some mechanism other than its gene product). Toxic proteins are distinguishable from simple chemical toxins (e.g., heavy metals and the like) by their antigenicity and higher molecular weight

In one embodiment, the toxic transgene comprises a gene encoding for an amyloidic protein, for example beta amyloid peptide, prion protein variants, transthyretin variants, gelsolin variants, cystatin variants, lysozyme variants and the like. In another embodiment, the transgene encodes a protein containing polyglutamine resulting from triplet-repeat expansion such as huntingtin (a protein that has been implicated in Huntington's Chorea). ataxin-1 or ataxin-2. Alternatively, proteins associated with inherent amyelotopic lateral sclerosis (ALS) for example, superoxide dismutase 1 variants and over-expressed neurofilament protein, can be used. It will be understood that the transgene can encode for an entire toxic protein or, alternatively, a functional (i.e. toxic) fragment.

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Transgenic Animals

In one aspect, the present invention includes a dually transgenic non-human animal. In one embodiment, the development of these dually transgenic animals involves the production of two *C. elegans* lines with a single transgene in each line, designated as CL2005 and CL2070. *C. elegans* was chosen due to the observation that the toxic effect of

 β -amyloid has been reported to occur in these animals, in addition to humans. Line CL2005 exhibited muscle-specific expression of human β peptide, while line CL2070 exhibited stress-inducible expression of GFP. GFP was incorporated into the present invention as it provides a marker (*i.e.*, "reporter") that is readily visible in living worms. Thus, dually transgenic animals both express β -peptide in their muscle cells and under appropriate conditions for observation (*i.e.*, fluorescence microscopy), exhibit green fluorescence in these cells.

Mating of these lines resulted in the production of dually transgenic animals, in which the presence of β -amyloid was easy to detect, due to the fact that the dually transgenic animals expressing β -amyloid glowed green. Thus, the present invention provides an easily detectable method for the specific expression of β -peptide in dually transgenic animals. This ease of detection provides great advantages for the development of methods to analyze the effects of β -amyloid in vivo. In addition, because the dually transgenic animals are not killed in order to detect the presence of β -amyloid, as would be required if other reporters (e.g., lacZ or luciferase) were used, the animals may be useful for screening compounds for their effects on β -amyloid in an in vivo situation. Thus, the use of an reporter which is detectable in vivo provides significant advantages over currently available methods.

Other Advantages of Transgenic C. elegans

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The present invention also includes methods of screening compounds for their ability to prevent or inhibit toxicity due to the toxic transgene. In these drug screening embodiments, the methods and animals described above can be used to identify potential therapies. For example, in assays to develop compounds to block the expression of β -amyloid or counter its toxic effects, transgenic worms (e.g., green fluorescing worms with muscle-specific β -peptide expression), would be observed for their loss of fluorescence. In assays where the compound is effective in countering the expression of β -peptide or effects of β -peptide, the animals would no longer glow green. It is easy to visually screen and quantify (using commercially available equipment) for animals that have lost the ability to glow, and then use them to detail the interaction between the compound and the toxic

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transgene. The present invention provides a fast and efficient screening system, for PCT/US97/23819 example by using commercially available equipment for assaying multiple compounds at

Thus, the transgenic animals of the present invention expressing a detectable reporter gene triggered by a toxic transgene provide animal models for human diseases. 5 For instance, a reporter gene (such as GFP) induced by expression of β-peptide provides an animal model for human Alzheimer's disease. It is also not intended that the present invention be limited to animals expressing β -peptide in conjunction with GFP. It is contemplated that expression of other proteins associated with dementia and/or

Alzheimer's or other diseases would also find use in conjunction with GFP in C. elegans as 10 well. For example, it is contemplated that proteins (including mutated versions of proteins) such as the presenilins associated with Alzheimer's disease (See e.g., J. Marx (1996) Science 274:1838-1840) will be used in conjunction with GFP and full-length APP in dually transgenic C elegans. Other toxic proteins which provide animal models for 15 other human diseases are described herein.

These transgenic animals also provide a means to screen compounds for their ability to decrease or eliminate the toxicity. Examples of compounds suitable for testing using the transgenic animals of the present invention include, but are not limited to, Congo Red, tumor necrosis factor (TNF), estrogen, tacrine (9-amino-1,2,3,4-tetrahydroacridine), dihydroepiandrosterone (DHEA). compounds that inhibit ApoE4. and others. commercially available from suppliers such as Sigma. Compounds to be tested for antitoxic activity are administered to the same number of dually transgenic animals (e.g., generated using the methods described in the Examples, below) from the control group and the treatment group, and the presence or absence of reporter gene used as a measure of efficacy.

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The compounds being tested can be administered using any suitable route (e.g., oral, parenteral, controlled-release transdermal methods, and implants, etc.). In one preferred route, the compounds to be tested are suspended in the growth media provided to the worms. Generally speaking, the route of administration will depend on the stability of the compound, the susceptibility of the compound to "first pass" metabolism, the

concentration needed to achieve a therapeutic effect, and the like. Following initial screening, a compound that appears promising (i.e., which increases the number of worms which display reduced β -peptide toxicity) is further evaluated by administering various concentrations of the compound to additional transgenic animals in order to determine an approximate therapeutic dosing range.

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Another screening method involves the crossing of the transgenic worms of the present invention with other transgenic worms. The animals are observed after treatment, in the presence and absence of the test compound(s), with the effects on the toxic transgene being gauged either by crude survival or the presence/absence of the reporter gene. It is also contemplated that the methods of the present invention be modified so as to provide means to analyze disease-related proteins believed to have dominant toxic effects, by substituting appropriate sequences for the β -peptide sequences used as described in the Examples. For example, proteins such as transthyretin (known to be associated with familial amyloid polyneuropathy; See e.g., Christmason et al., (1991) FEBS 281:177-180; Genbank Accession #D00096), and variant superoxide dismutase (known to be associated with familial amyotrophic lateral sclerosis [Lou Gehrig's disease]), prion proteins, A4 amyloid protein (See e.g., Ponte et al., (1988) Nature 331:525-527; Salbaum et al., U.S. Patent No. 5,151,508, herein incorporated by reference), APP (See e.g., Kitaguchi et al., (1988) Nature 331:530-532; Sata et al., EP Appln. 94117512.7; Scott et al., WO 9412627; Wadsworth et al., WO 9314200; Gearhart et al., WO 9423049; and Neve et al., WO 9302189, all of which are herein incorporated by reference), other amyloidic proteins (e.g., variant lysozymes and amylin peptide), and other proteins associated with neuronal degeneration (See e.g., U.S. Patent No. 5,196,333), will be used in the methods of the present invention by substituting the appropriate nucleic acid sequences encoding the protein of interest for the β -peptide described in Example 1. It is also contemplated that animals transgenic for dual proteins (e.g., \beta-peptide in combination with another protein. such as transthyretin) will be used. Thus, the present invention provides the means to analyze the effects of numerous genes and proteins in vivo.

Furthermore, the transgenic animals of the present invention provide distinct advantages over other transgenic animals currently used to analyze diseases such as

Alzheimer's disease. Due to the short gestation period of C. elegans, transgenic animals can be produced much more rapidly than when mammals, such as mice are used. For example, transgenic mice overexpressing a 695 amino acid isoform of β -amyloid precursor did not show learning and memory impairment until they were 9-10 months of age (Hsiao et al., Science 274:99-102 [1996]). In contrast, the transgenic animals of the present invention express β -peptide and may be manipulated at a very early age, even in embryonic stages.

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In addition, because the transgenic animals are easy to select based on the presence of the green fluorescence, invasive procedures such as surgery, necessary to analyze the effects of the toxic transgene in other animals (e.g., mice) are avoided.

Finally, the apparent toxicity of β-peptide in transgenic animals was found to be temperature-dependent. Transgenic animals maintained at 25.5°C were significantly more sick than those maintained at 16°C. The animals maintained at the higher temperature became paralyzed more quickly, failed to eat or grow, failed to show normal egg-laying, and many died before reaching adulthood. However, wild-type animals grow well at either of these temperatures. The CL2005 parental line was temperature-sensitive for viability, as it was not possible to propagate this line at the elevated temperature. In the CL2070 parental line, 25.5°C maintenance was not sufficient to induce the hsp/GFP construct. The hsp/GFP response was similarly temperature-dependent. It was difficult to detect GFP when the dual transgenic animals were raised at 16°C, but GFP induction was dramatic when the animals were raised at 25.5°C.

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These temperature dependency observations were exploited by the propagation of animals at 16°C, and then upshifting them to 25.5°C, when GFP induction was desired. This effect also has potential benefits in the analysis of compounds, as the animals can be pre-incubated in the presence of drugs or other compounds at the lower temperature before the upshift to the higher temperature, in order to ensure that the presumed protective effect of the drug was in place, prior to the strong induction of β -peptide toxicity. Importantly, these assays may be completed within one day and are suitable for rapid methods (e.g., the use of a microtiter format and a plate fluorimeter), so that literally thousands of compounds may be tested simultaneously.

In sum, the present invention provides methods and compositions useful as animal models for disease, as well as providing methods and compositions for disease therapy and prevention. The animal testing may be supplemented and confirmed by testing on human subjects. However, the transgenic animals of the present invention allow the testing of a large number of compounds, both various methods, including those known in the art.

DEFINITIONS

To facilitate understanding of the invention, a number of terms are defined below.

The term "non-human animal" includes vertebrates such as rodents, arthropods, insects (e.g., Diptera), fish (e.g., zebrafish), non-human primates, ovines, bovines, ruminants, lagomorphs, porcines, caprines, equines, canines, felines, aves, etc. The non-human animal can be used at any stage in its development, for example oocyte, fetal, larval, pupal stages or the like. Preferred non-human animals are those that are transparent, such as certain nematodes, oocytes, larvae and fish.

As used herein, the term "transparent" is used in references to animals through which light will be transmitted. However, it is not intended that the amount of light transmittance be limited to any particular amount. For example, an animal is transparent so long as at least some light may be transmitted through its body. Non-limiting examples

of transparent animals are larval stages of some animals (e.g., flies) and oocytes (e.g.,

Xenopus oocytes). Particularly preferred non-human animals are selected from the nematodes (i.e., any animal in the Class Nematoda), most preferably Caenorhabditis elegans. However, it is contemplated that other transparent animals, such as zebrafish will be useful in the present invention. For example, transgenic zebrafish have been produced (See e.g., Lin et al., PCT Publ. WO9603034; incorporated herein by reference).

The "non-human animals having a genetically engineered genotype" of the invention are preferably produced by experimental manipulation of the genome of the germline of the non-human animal. These genetically engineered non-human animals may be produced by several methods including the introduction of a "transgene" comprising nucleic acid (usually DNA) into an embryonal target cell or integration into a chromosome of the somatic and/or germ line cells of a non-human animal by way of human

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intervention, such as by the methods described herein. The process by which a DNA molecule becomes stably incorporated into another genome is referred to as "stable integration." Non-human animals which contain a transgene are referred to as "transgenic non-human animals". A transgenic animal is an animal whose genome has been altered by the introduction of one or more transgenes.

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The term "transgene" as used herein refers to a foreign gene that is placed into an organism by introducing the foreign gene into gonadal cells, embryonic cells, newly fertilized eggs, or early embryos. The term "foreign gene" refers to any nucleic acid (e.g., manipulations and may include gene sequences found in that animal by experimental introduced gene contains some modification relative to the naturally-occurring gene. A preferred foreign gene is the β-amyloid gene (e.g., β-peptide), or fragments thereof. A for example, the toxic transgene may produce a gene product (i.e., protein) which is toxic to the cell or organism. Alternatively, the transgene may disrupt other cellular proteins, or act as a DNA binding site without producing a protein

As used herein, the term "toxicity" refers to the production of toxic effects by any compound or substance. For example, while it is not necessary to the understanding and use of the present invention, β -peptide toxicity may arise due to the accumulation of β -peptide molecules. There are numerous manifestations of toxicity that may occur. For example, the toxicity associated with β -peptide may be manifested as negative effects on muscle cells of dually transgenic animals.

The terms "promoter element," "promoter," or "promoter sequence" as used herein, refer to a DNA sequence that is located at the 5' end of (i.e., precedes) a gene in a DNA polymer and provides a site for initiation of the transcription of the gene into mRNA. An instance binding of a transcription factor, stress heat or the like

The term "reporter gene" as used herein refers to genes that encode proteins or other compounds that can be detected by a variety of methods. These genes "report" the occurrence of successful introduction and expression of sequences such as transgenes.

Non-limiting examples of reporter genes include antibiotic resistance genes, genes encoding enzymes and genes encoding other detectable proteins. Expression of the reporter gene is detected using methods known in the art. In a preferred embodiment, the reporter gene is a GFP gene, although lacZ β -galactosidase gene, or any other reporter system may be used to detect the successful production of transgenic animals. In particularly preferred embodiments, the reporter is a compound or protein which may be present or expressed within living animals. That is, it is not necessary to sacrifice the animal in order to detect the presence of the reporter. The type of the reporter gene system used is not critical to the invention, and it is contemplated that any system suitable for use with the transgenic animals of the present invention will be used.

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As used herein, the terms "dual trangenic" and "dually transgenic" refer to animals or cells in which more than one transgene have been introduced. For example, the term is used in reference to cells which contain the sequences encoding β -peptide, and sequences encoding a reporter (e.g., GFP). However, it is not intended that the number of transgenes in the dually transgenic animals of the present invention be limited to two. For example, the transgenic animals may also contain another one or more marker genes (e.g., rol-6 sequences), in addition to β -peptide and reporter sequences. In the embodiments in which rol-6(su-1006), expression of the marker gene is accomplished by observing the living animals for the rolling behavior that results due to the expression of the abnormal collagen gene. In contrast, wild-type animals exhibit normal locomotion and do not exhibit this rolling behavior, as their collagen is normal.

The transgenic animals of the present invention are preferentially generated by introduction of the targeting vectors into gonad cells. Transgenes can be efficiently introduced into the cells by DNA transfection using a variety of methods known to the art, including electroporation, calcium phosphate co-precipitation, protoplast or spheroplast fusion, lipofection, and DEAE-dextran-mediated transfection. Transgenes may also be introduced into cells by retrovirus-mediated transduction or by micro-injection. In one preferred embodiment, the transgenes are injected into gonads of *C. elegans* as described by Mello *et al.* (1991) *EMBO J.*. **10**:3959-3970. Alternative methods for the generation of transgenic animals containing an altered gene are known to the art. For example,

embryonal cells at various developmental stages can be used to introduce transgenes for the production of transgenic animals. Different methods are used depending on the stage of development of the embryonal cell.

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A wide variety of labels and conjugation techniques are known by those skilled in the art and can be used in various nucleic acid and amino acid assays. Means for producing labeled hybridization or PCR probes for detecting related sequences include oligolabeling, nick translation, and end-labeling or PCR amplification using a labeled nucleotide. Suitable reporter molecules or labels include those radionuclides, enzymes, fluorescent, chemiluminescent, or chromogenic agents as well as substrates, cofactors, inhibitors, magnetic particles and the like. The term may also be used in reference to proteins. For example, a variety of protocols which employ polyclonal or monoclonal antibodies specific for the β-peptide protein product are known in the art (See, the Examples). These antibodies can be used as markers for the expression of proteins. Examples include enzyme-linked immunosorbent assay (ELISA), radioimmunoassay (RIA) and fluorescent activated cell sorting (FACS).

The term "gene" refers to a DNA sequence that comprises coding sequences necessary for the production of a polypeptide or precursor. In some instances, a gene can also include control sequences. The polypeptide can be encoded by a full length coding sequence or by any portion of the coding sequence so long as the desired enzymatic activity is retained.

The term "gene of interest" refers to any gene, the manipulation of which may be deemed desirable for any reason, by one of ordinary skill in the art.

The term "wild-type" refers to a gene or gene product which has the characteristics of that gene or gene product when isolated from a naturally occurring source. A wild-type gene is that which is most frequently observed in a population and is thus arbitrarily designed the "normal" or "wild-type" form of the gene. In contrast, the term "modified" or "mutant" refers to a gene or gene product which displays modifications in sequence and or functional properties (i.e., altered characteristics) when compared to the wild-type gene or gene product. It is noted that naturally-occurring mutants can be isolated: these are

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tified by the fact that they have altered characteristics when compared to the wild-type or gene product.

The terms "targeting vector" or "targeting construct" refer to oligonucleotide inces comprising a gene of interest flanked on either side by regulatory sequences. ably, the targeting vector is capable of homologous recombination such that the gene rest is integrated by recombination.

As used herein, the terms "vector" and "vehicle" are used interchangeably in ice to nucleic acid molecules that transfer DNA segment(s) from one cell to another. The terms "expression vector" or "expression cassette" as used herein, refers to a inant DNA molecule containing a desired coding sequence and appropriate nucleic quences necessary for the expression of the operably linked coding sequence in a ir host organism. Nucleic acid sequences necessary for expression in prokaryotes include a promoter, an operator (optional), and a ribosome binding site, often along it sequences. Eukaryotic cells are known to utilize promoters, enhancers, and in and polyadenylation signals.

ne terms "in operable combination." "in operable order," and "operably linked" as n, refer to the linkage of nucleic acid sequences in such a manner that a nucleic rule capable of directing the transcription of a given gene and/or the synthesis of rotein molecule is produced. The term also refers to the linkage of amino acid n such a manner so that a functional protein is produced.

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term "tissue-type specific" as it applies to a promoter, refers to a promoter that f selectively directing expression of a gene in a specific tissue. Similarly, the ne-specific promoter in the method of the present invention does not require cificity. In general, the requisite specificity is found where a plurality (or, bly, a majority) of cells in one tissue type express a gene of interest, while negative than 80%, and preferably greater than 90%, and more preferably 5%), of the cells in other tissue types do not. In one embodiment of the ion, the strong muscle promoter unc54 was used (See, Example 1).

In the term "portion" when in reference to a gene refers to fragments of fragments may range in size from a few nucleotides to the entire gene

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sequence minus one nucleotide. Thus, "an oligonucleotide comprising at least a portion of a gene" may comprise fragments of the gene or the entire gene.

As used herein, the term "substantially purified" refers to molecules, either nucleic or amino acid sequences, that are removed from their natural environment, isolated or separated, and are at least 60% free, preferably 75% free, and most preferably 90% free from other components with which they are naturally associated. An "isolated polynucleotide" is therefore a substantially purified polynucleotide.

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As used herein, the term "polymerase chain reaction" ("PCR") refers to the method of K.B. Mullis disclosed in U.S. Patent Nos. 4,683,195, 4.683,202 and 4,965,188, all of which are hereby incorporated by reference, which describe a method for increasing the 10 concentration of a segment of a target sequence in a mixture of genomic DNA without cloning or purification. This process for amplifying the target sequence consists of introducing a large excess of two oligonucleotide primers to the DNA mixture containing the desired target sequence, followed by a precise sequence of thermal cycling in the presence of a DNA polymerase. The two primers are complementary to their respective 15 strands of the double stranded target sequence. To effect amplification, the mixture is denatured and the primers then annealed to their complementary sequences within the target molecule. Following annealing, the primers are extended with a polymerase so as to form a new pair of complementary strands. The steps of denaturation, primer annealing and polymerase extension can be repeated many times (i.e., denaturation, annealing and 20 extension constitute one "cycle": there can be numerous "cycles") to obtain a high concentration of an amplified segment of the desired target sequence. The length of the amplified segment of the desired target sequence is determined by the relative positions of the primers with respect to each other, and therefore, this length is a controllable parameter. By virtue of the repeating aspect of the process, the method is referred to as the 25 "polymerase chain reaction" (hereinafter "PCR"). Because the desired amplified segments of the target sequence become the predominant sequences (in terms of concentration) in the mixture, they are said to be "PCR amplified."

With PCR, it is possible to amplify a single copy of a specific target sequence in genomic DNA to a level detectable by several different methodologies (e.g., hybridization

with a labeled probe; incorporation of biotinylated primers followed by avidin-enzyme conjugate detection; and/or incorporation of ³²P-labeled deoxynucleotide triphosphates, such as dCTP or dATP, into the amplified segment). In addition to genomic DNA, any oligonucleotide sequence can be amplified with the appropriate set of primer molecules. In particular, the amplified segments created by the PCR process itself are, themselves, efficient templates for subsequent PCR amplifications. Amplified target sequences may be used to obtain segments of DNA (e.g., genes) for the construction of targeting vectors, transgenes, etc.

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As used herein, the terms "PCR product" and "amplification product" refer to the resultant mixture of compounds after two or more cycles of the PCR steps of denaturation, annealing and extension are complete. These terms encompass the case where there has been amplification of one or more segments of one or more target sequences.

The term "Central Nervous System" refers to the "spinal cord" and the "brain." The spinal cord comprises white areas and grey areas. The grey area contains nerve cell bodies, whereas the white area is essentially comprised of myelinated nerves. The brain, which is also known as the encephalon, is that portion of the cerebrospinal axis which is contained in the cavity of the cranium. The brain comprises the two cerebral hemispheres, the interbrain, the mid-brain, the pons Varolli and cerebellum, and the medulla oblongata. The two hemispheres together with the parts derived from the thalamencephalon form the forebrain. The two cerebral hemispheres are separated by the longitudinal fissure and also comprise the bylvian fissure, the fissure of Rolando, and the parieto-occipital fissure. The lobes on the external surface of the brain comprise the frontal lobe, the parietal lobe, the occipital lobe, and the temporal lobe. Placed along the middle line of the brain are, among others, the rostrum and peduncles of corpus callosum, lamina cinera, optic commissure and the pituitary body. On each side of the middle line lies the frontal lobe, olfactory lobe and the hemisphere of cerebellum.

The terms "neuron," "neural cell," and "nerve cell" are used interchangeably to refer to a cell which is located in the nervous system. Nerve cells are composed of the nerve cell body (perikaryon), one or more dendrites, and an axon. Neurons can be classified according to the number of processes originating from the cell body. Thus,

unipolar neurons have a single process, bipolar neurons have one axon and one dendrite, while multipolar neurons (which are the most common) comprise more than two processes. The term "neuron" comprises cholinergic neurons and sensory neurons. As used herein, the term "cholinergic neuron" means a neuron in the Central Nervous System (CNS) and in the Peripheral Nervous System (PNS) whose neurotransmitter is acetylcholine. As used herein, the term "sensory neuron" includes a neuron which is responsive to environmental cues (e.g., temperature and movement) from, for example, the skin, muscle and joints of a mammal.

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The term "nerve" refers to two or more neurons arranged in linear sequence such that the axon of one neuron establishes a structural and functional link with the dendrite of a second neuron to form a "synapse."

The term "compound" refers to any chemical entity, pharmaceutical, drug, and the like that can be used to treat or prevent a disease, illness, sickness, or disorder of bodily function. Compounds comprise both known and potential therapeutic compounds. A compound can be determined to be therapeutic by testing using the testing methods of the present invention (i.e., a "test compound"). A "known therapeutic compound" refers to a therapeutic compound that has been shown (e.g., through animal trials or prior experience with administration to humans) to be effective in such treatment or prevention. In other words, a known therapeutic compound is not limited to a compound efficacious in the treatment or prevention of neural related disorders.

A compound is said to be "in a form suitable for administration such that the compound is bioavailable in the blood of the animal" when the compound may be administered to an animal by any desired route (e.g., oral, intravenous, subcutaneous, intramuscular, etc.) and the compound or its active metabolites appears in the blood of the animal in an active form. Administration of a compound to a pregnant animal may result in delivery of bioavailable compound to the embryonic progeny of the animal.

The "wild-type β-amyloid" or "β-peptide" gene and gene product refers to the nucleotide and amino acid sequences provided in SEQ ID NOS:6 and 7, respectively. Those skilled in the art will be well aware that certain modifications of SEQ ID NOS:6 and 7 can be made which will not interfere with the production of a polypeptide having an

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activity indistinguishable from that of the wild-type β-amyloid; the present invention specifically contemplates these variant forms of β-amyloid. A "variant" of the β-peptide is defined as an amino acid sequence that differs by one or more amino acids. The variant may have "conservative" changes, wherein a substituted amino acid has similar structural or chemical properties (e.g., replacement of leucine with isoleucine). More rarely, a variant may have "nonconservative" changes, e.g., replacement of a glycine with a tryptophan. Similar minor variations may also include amino acid deletions or insertions (i.e., additions), or both. Guidance in determining which and how many amino acid residues may be substituted, inserted or deleted without abolishing biological or immunological activity may be found using computer programs well known in the art, for example, DNAStar software.

The term "an oligonucleotide sequence comprising at least a portion of a human β -amyloid gene" refers to a polynucleotide sequence (*i.e.*, a nucleic acid sequence) containing a nucleotide sequence derived from a human β -amyloid gene. This sequence may encode a portion or all of the β -amyloid protein; alternatively, this sequence may contain non-coding regions derived from the β -amyloid gene or a combination of coding and non-coding regions. The oligonucleotide may be RNA or DNA and may be of genomic or synthetic origin.

As used herein the term "portion" when in reference to a gene refers to fragments of that gene. The fragments may range in size from 10 nucleotides to the entire gene sequence minus one nucleotide. Thus, "an oligonucleotide comprising at least a portion of a gene" may comprise fragments of the gene or the entire gene.

An animal whose genome comprises a "heterologous marker gene" is an animal whose genome contains a marker gene not naturally found in the animal's genome. In one preferred embodiment of the present invention, the heterologous marker gene is a mutant collagen gene, such as the rol-6(su-1006) gene. However, it is intended that other marker genes will be used with success in the present invention, including other mutant collagen genes, as well as other marker genes commonly known to those in the art.

As used herein, the term "diagnostic assay" refers to methods for the diagnosis of disease, illness, and/or pathology. It is intended that the term encompass any methods for

diagnosis, including, but not limited to assays based on immunoreactivity (e.g., radioimmunoassays, fluorescence immunoassays, enzyme immunoassays), histochemistry, dye retention or binding (e.g., fixing of dyes such as Congo Red), nucleic acid based diagnostic methods (e.g., identification of nucleic acid sequences associated with disease or pathology), etc.

EXPERIMENTAL

The following examples are provided in order to demonstrate and further illustrate certain preferred embodiments and aspects of the present invention and are not to be construed as limiting the scope thereof.

In the experimental disclosure which follows, the following abbreviations apply: eq (equivalents); M (Molar); μM (micromolar); N (Normal); mol (moles); mmol (millimoles); μmol (micromoles); nmol (nanomoles); g (grams); mg (milligrams); μg (micrograms); ng (nanograms); l or L (liters); ml (milliliters); µl (microliters); cm (centimeters); mm (millimeters); μm (micrometers); nm (nanometers); °C (degrees Centigrade); U (units), mU (milliunits); min. (minutes); sec. (seconds); % (percent); kb (kilobase); bp (base pair); PCR (polymerase chain reaction); β-(1-42) amino acids 1-42 of β-amyloid peptide; Tris (tris(hydroxymethyl)-aminomethane); BSA (bovine serum albumin); Fisher (Fisher Scientific, Pittsburgh, PA); Sigma (Sigma Chemical Co., St. Louis, MO.); Promega (Promega, Corp., Madison, WI); Perkin-Elmer (Perkin-Elmer/Applied Biosystems, Foster City, CA); Senetek (Senetek, PLC, Maryland Heights, MO); Boehringer Mannheim (Boehringer Mannheim, Corp., Indianapolis, IN); Stratagene (Stratagene, Inc., La Jolla, CA); and NEB (New England Biolabs, Beverly, MA). Unless otherwise indicated, the restriction enzymes used in these Examples were obtained from NEB. C. elegans is available from the Caenorhabditis Genetics Center, at the University of Minnesota, St. Paul, MN.

EXAMPLE 1

Assembly of the β -(1-42) Minigene

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1-42) minigene used in subsequent experiments (See. ic C. elegans engineered to express amyloidic human escribed below, these animals contained constructs in 54 promoter/enhancer of C. elegans drove the expression of lerived from human β-amyloid cDNA clones, as described ıman β-amyloid peptide in transgenic Caenorhabditis , 92:9368-9372 [1995]). ng the 42 amino acid β-amyloid peptide derived from in cDNA was assembled. The artificial signal peptide 52.81 was amplified under standard conditions, using "GGCCAAAGGACCC-3")(SEQ ID NO:1), and "SP-down" CCAGCAAGAT-3')(SEQ ID NO: 2), cleaved with NheI nd inserted between the unique Nhel and Kpnl sites of : construct "pCL2." This process resulted in a res. such that the signal peptide cleavage site, as predicted by von Heijne, Nucl. Acids Res., 14:4683-4690 [1986]), Jly-Thr dipeptide encoded by the KpnI site. ding amino acids 1-42 of β-amyloid, and which contained olified using standard methods, from human \(\beta\)-amyloid 4T4B (P. Ponte et al., Nature 331:525-527 [1988]), by GGGGTACCGATGCAGAATTCCGACATGA-3') (SEO 5'-CCCGAGCTCACGCTATGACAACACCGCCAAcation product was cleaved with KpnI and SacI, and il and Sac I sites of pCL2, to generate "pCL3." 12) minigene fragment was removed from this plasmid by d inserted between the unique Nhel and Sacl sites of The sequence of the β -(1-42) minigene was confirmed by coding strand only, by techniques known in the art. The):5), is shown in Figure 2. A graphic map of pCL12 is hows the DNA and amino acid sequences (SEQ ID NOS:

6 and 7, respectively) of pCL12 from nucleotide 1071 through 1253 (i.e., the $\beta(1-42)$ nucleic acid and amino acid sequences) present in the construct.

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EXAMPLE 2

Construction of the hsp/GFP Reporter

imple, the expression vector ("pCL25") containing the hsp/GFP reporter expression of the GFP-TT gene was constructed. This vector was used in ments (See, Example 3) to produce transgenic C. elegans was 2-TT is a modified form of GFP that contains Ser65Thr and Ile167Thr relative to the sequence of the wild-type GFP). The substitutions present ce a protein which results in the production of a much brighter the wild-type GFP.

as constructed as described below. A 431 bp *Hin*dIII-*Bam*HI fragment p16-2 promoter was excised from the plasmid "pPD49.78." by digestion *Hin*dIII. This 431 bp *Hin*dIII-*Bam*HI fragment was inserted between the de #1) and *Bam*HI (nucleotide #31) sites of the GFP-TT gene in the TT," (available from Yishi Jin. at the University of California, Santa igase (Promega). The pGFP-TT plasmid contains the coding regions for into a Tu61 backbone. The DNA sequence of pGFP-TT (SEQ ID NO: 8) re 5. A schematic map of pGFP-TT is shown in Figure 6. in which unique are shown. The pGFP-TT plasmid was digested with *Hin*dIII and *Bam*HI, ragment containing the hsp16-2 promoter was inserted, in order to ss-inducible expression vector "pCL25." The DNA sequence of pCL25 and restriction map are shown in Figure 7. Figure 8 provides a schematic n. which unique restriction sites are shown.

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EXAMPLE 3

Construction of Transgenic Animals

In this Example, the transgenic parent animals were produced, with one line expressing the β(1-42) minigene (designated as "CL2005") and the other line expressing the hsp/GFP reporter (designated as "CL2070"). For both lines, the transgenes were introduced into *C. elegans* by gonad microinjection as known in the art and described by Mello *et al.* (Mello *et* EMBO J., 10:3959-3970 [1991]). Marker plasmid pRF4 containing Mello *et al.* (Mello *et* EMBO J., 10:3959-3970 [1991]). Marker plasmid pRF4 containing the gene (*rol*-6[su1006]) (SEQ ID NO:9)(pRF4 carries a 4 kb *Eco*RI fragment of *C. elegans* genomic DNA containing the *rol*-6[su1006] collagen gene in the Bluescribe vector [Stratagene]: *See.* Mello *et al.. supra*; and Kramer *et al..* Mol. Cell. Biol., 10:2081-2090 [Stratagene]: *See.* Mello *et al.. supra*; and Kramer *et al..* Mol. Cell. Biol., 10:2081-2090 [1990]) was coinjected with the constructs, at approximately 100 ng/μl for each plasmid. Into morphologically wild-type animals, and Roller transgenic progeny were recovered. The (*rol*-6[su1006]) gene (SEQ ID NO:9), contained within the pRF4 plasmid is a mutated *C. elegans* collagen gene, the expression of which produces the dominant, distinctive "Roller" phenotype. Figures 9 and 10 show the DNA and amino acid sequences of the *rol*-6 gene used in this Example, respectively.

Transmitting lines were established and maintained by selection for the Roller marker phenotype. Transgenic animals produced in this manner maintain the injected DNA as an extrachromosomal, multicopy array of variable mitotic and meiotic stability. Strains containing chromosomally integrated transgenes were recovered by irradiation of lines containing extrachromosomal transgenic arrays with 7000 rad (1 rad = 0.01 Gy) of lines containing extrachromosomal transgenic arrays with 7000 rad (1 rad = 0.01 Gy) of lines arrays from a Cesium-66 source. Progeny of irradiated animals were then screen for 100% transmittance of the marker transgene. The transgenes in both the CL2005 and CL2070 lines were chromosomally integrated, and were 100% stable.

Transgenic (i.e., as indicated by the Roller phenotype) animals produce both transgenic (i.e., Roller) and non-transgenic (i.e., non-Roller) progeny. These non-transgenic progeny were found to serve as good internal controls for phenotypic and immunohistochemical comparisons. The expression of GFP in the dual transgenic animals immunohistochemical comparisons. The expression of animals from 16 to 25.5°C, at all can be detected in less than 24 hours after the upshift of animals from 16 to 25.5°C, at all

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stages of development, from late embryonic to adult. Expression of GFP resulted in the production of green fluorescence in muscle cells; intense tissue-specific expression of GFP was observed. The fluorescence can be observed using compound or dissecting epifluoroescence microscopy (*i.e.*, with standard fluorescein excitation and emission filters). It is also contemplated that the fluorescence is observable by use of fluorimeters and cell sorters.

EXAMPLE 4

Immunohistochemistry of Transgenic Animals

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In this Example, immunohistochemistry was used to confirm the transmittance of the chimeric constructs in large populations of putative integrated lines.

As described by Link (Link, 1995, supra), whole mount specimens were prepared by fixing animals in 4% paraformaldehyde and permeabilizing them with 2-mercaptoethanol collagenase as known in the art and described by Link et al. (Link et al., Genetics 131:867-881 [1992]). Three antibodies were used (polyclonal rabbit anti-β peptide antibody, Boehringer Mannheim; mouse monoclonal antibody Ab 4G8, available from Senetek: and monoclonal 4.1, a gift from B. Cordell at Scios Nova). The monoclonal 4.1 recognizes residues 8-15 of β-peptide.

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As was observed by Link (Link, 1995, supra) animals transgenic for the unc-54/β(1-42) minigene construct contained muscle-specific deposits of anti-β peptide immunoreactivity. Non anti-β-immunoreactivity was observed in control animals (i.e., non-transgenic, wild-type C. elegans tested concurrently). Although C. elegans is reported to contain a homolog of the β-amyloid precursor protein gene (See, Daigle and Li, Proc. Natl. Acad. Sci., 90:2045-2049 [1993]), this sequence does not contain an apparent β-peptide domain. Thus, it would not be expected, nor did it show, cross-reactivity with the anti-β antibody used in these experiments. For dually transgenic animals, immunoreactive deposits accumulated in the body wall of the animals.

Next. to determine whether the immunoreactive deposits observed in the unc-54/ β -(1-42) strains displayed the tinctural properties of classic insoluble β -amyloid, transgenic

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strains were fixed and stained with thioflavin S, a fluorescent amyloid-specific dye, as known in the art, and described by Guntern and Bouras (R. Guntern and C. Bouras, Experientia 48:-10 [1992]). Thioflavin S-reactive deposits were found in all strains containing the $unc-54/\beta-(1-42)$ minigene constructs, but not in control wild-type animals.

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Whole mounts of fixed dual transgenic animals stained with coumarin-phallacidin (a muscle-specific probe) and anti- β peptide antibody. Muscle-specific β peptide deposits were observed, and showed a qualitative correlation with the level of GFP expression in these muscle cells (*i.e.*, cells that contained more β -peptide deposits were more green than cells with less β -peptide deposits).

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In addition, Congo Red and Chrysamine G, two dyes known to interact with β -amyloid and have been reported to interfere with its aggregation are tested on the dually transgenic animals. Anti-oxidants (e.g., vitamin E and ascorbate) are also tested. In these experiments, interference with aggregation of β -peptide, or interaction with β -amyloid are observed.

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All publications and patents mentioned in the above specification are herein incorporated by reference. Various modifications and variations of the described method and system of the invention will be apparent to those skilled in the art without departing from the scope and spirit of the invention. Although the invention has been described in connection with specific preferred embodiments, it should be understood that the invention as claimed should not be unduly limited to such specific embodiments. Indeed, various modifications of the described modes for carrying out the invention which are obvious to those skilled in molecular biology or related fields are intended to be within the scope of the following claims.

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CLAIMS

What is claimed is:

5 1. A transparent animal comprising:

- (a) a toxic transgene;
- (b) an inducible promotor operably linked to a reporter gene, wherein the toxic transgene induces the promotor and wherein the expression of the reporter gene is detectable *in vivo*.

2. The transparent animal of claim 1, wherein the inducible promoter operably linked to the reporter gene is stably integrated into the genome of the animal.

- 3. The transparent animal of claim 2, wherein the inducible promotor is induced by disruption of cellular function.
 - 4. The transparent animal of claim 2, wherein the inducible promoter is a heat shock promoter.
 - 5. The transparent animal of claim 2, wherein the toxic transgene is β -peptide.
 - 6. The transparent animal of claim 2, wherein said transparent animal is selected from the class Nematoda.
- 7. The transparent animal of Claim 6, wherein said transparent animal is Caenorhabditis elegans.

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	8.	The transparent animal of Claim 2, wherein said genome comprises SEQ ID
NO:5 a	and SEC) ID NO:8.

- The transparent animal of Claim 2, wherein said reporter gene is green fluorescence protein (GFP).
 - 10. The transparent animal of claim 1, further comprising a heterologous gene marker.
 - 11. A method for producing a dually transgenic non-human animal comprising:
 - a) providing:
 - i) a first and second non-human animal; and
 - ii) a first toxic transgene; and
 - iii) a second transgene, comprising a reporter;
 - b) introducing said first toxic transgene into the genome of said first non-human animal to produce a first transgenic animal, and introducing said second transgene into the genome of said second non-human animal to produce a second transgenic animal; and
 - c) mating said first transgenic animal with said second transgenic animal to produce a dually transgenic animal, wherein said toxic transgene and said reporter are expressed.
 - 12. A dually transgenic non-human animal produced according to the method of claim 11, wherein the toxic transgene is β -peptide.
 - 13. A dually transgenic non-human animal produced according to the method of claim 11, wherein said non-human animal is transparent:

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The dually transgenic non-human animal of claim 13, wherein said non-14. human animal is a nematode. The dually transgenic non-human animal of claim 14, wherein said 15. nematode is Caenorhabditis elegans The method of claim 11, wherein said reporter is green fluorescent protein. 16. The dually transgenic non-human animal produced according to the method 17. of claim 11, wherein said first transgene comprises pCL25. The dually transgenic non-human animal produced according to the method 18. of claim 11, wherein said second transgene comprises pCL12. The dually transgenic non-human animal produced according to the method 19. of claim 11, further comprising a heterologous marker gene. A method for testing compounds for toxicity, comprising: 20. a) providing: i) a dually transgenic non-human animal expressing a toxic transgene and reporter gene operably linked to a promoter inducible by the toxic transgene; ii) a composition comprising a test compound in a form suitable for administration such that said compound is bioavailable in the cells of said nonhuman animal; and

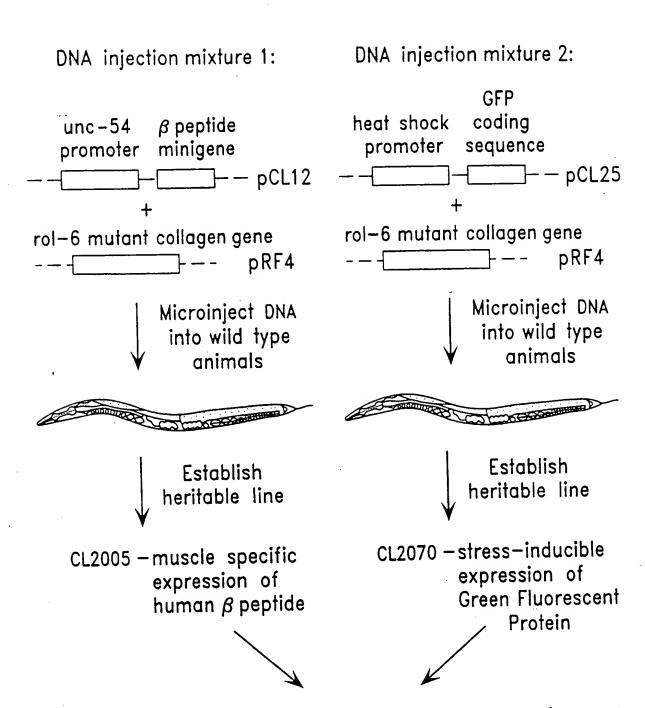
b) administering said test compound to said non-human animal.

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21. The method according to claim 20, wherein the toxic transgene is β -peptide and the reporter gene is GFP.

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- The method of Claim 21, further comprising c) measuring a reduction in the fluorescence of said non-human animal and thereby identifying a compound as therapeutic.
- The method of Claim 20, wherein said compounds inactivate said β-peptide
 expressed by said dually transgenic animal.



Mate lines together, recover dual transgenic animals by classic genetic techniques

Fig. 1

Fig. 2A
Fig. 2B
Fig. 2C
Fig. 2D
Fig. 2E
Fig. 2F
Fig. 2G
Fig. 2H
Fig. 2I
Fig. 2J
Fig. 2K
Fig. 2L
Fig. 2M
Fig. 2N
Fig. 20
Fig. 2P
Fig. 2Q
Fig. 2R
Fig. 2S
Fig. 2T

> Fig. 2

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		10/68			2H
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AACGACCTACACCGA/	MSP I HPG II HPG TI CCGTAA	CCTGTCGGGTTT	Hae Fnu4H Bs+U I	
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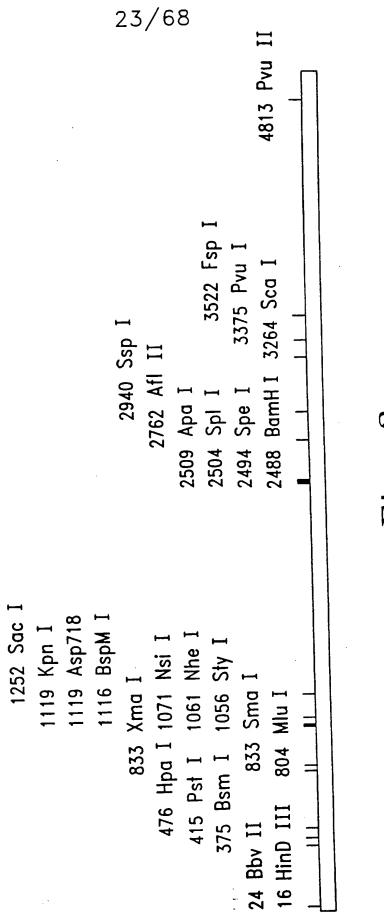


Fig. 3

Fig. 4

<u>a</u> | a 909 ACC GAT asb a l a phe thr GCA GGT gly 110 phe GTG <u>a | a</u> lys leu val CAA AAA TTG CCA pro <u>a | a</u>) nel gln 51 31 phe 1101 / ATC TTT 1161 / his CAT CAT 1221 his glu val phe GAA GTT bhe . 110 **CTG** ala leu TCA GGA TAT tyr CCA gly CTG l eu ser GAC 116 l e u asb CAT 1 AAG GTT / d / his 1 y s 2 1 CGA arg 41 glu phe 1191 / ATG CAT met his GAA TTC

GTC Va | gly val GTG GGC GGT GTT gly leu met val gly ATG CIC GGA TCA AAC AAA GGT GCA ATC ATT <u>.</u> <u>..</u> ala gly lys asu ser CGT GAT GTG

asp val gly 1251 / 61

TGA

Fig. 5A Fig. 5B Fig. 5C Fig. 5D Fig. 5E Fig. 5F
Fig. 5C Fig. 5D Fig. 5E
Fig. 5D Fig. 5E
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Fig. 5B
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!	560	→ Thr 167 AC 640 TG	720	C C	Fig.
	bo II AAGATGGAAAC TTCTACCTTTG	ACTATAACTCACACAATGTACATCATGCAGACAAACAAAGAATGGAATCAAAGTTAACTTCAAAAGTATGAAAGTTTAACTTCAAAAGTTTAACTTTAACTTTAAAAGTTAAAAGTTTAAAAGTAAAAGAATGAAAGAATTAAAAAA	Mae I CAACTAGCAGACCATTATCAACAAATACTCCAATTGGCG GTTGATCGTCTGGTAATAGTTGTTTTATGAGGTTAACCGC	Sau3A I Mbo I Dpn I Atw I I Atw I I Atw I I Atw I I I Atw I I Atw I I Atw I I I Atw I	AGACAACCATTACCTGTCCACACAATCTGCCCTTTCGAAAGATCCCAACGAAAGGAGGACGAGGAGGAGGAGGAGGAGGAGGA

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880	Fig.
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	31	/68		5F
1040	1120 1120 20	1200	1280	Fig.
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1840	1920	de I 2000 000	Fig.
ACAGCTTGTCTGTAAGCGGATGCCGGGAGCAAGCCCGTCAGGGCGGTGTTGGCGGGTGTCGGGGTGTGAGCCCCGACACCGGGTGTTGGCGGGTGTCGGGGGTGTTGGCGGGTGTTGGCGGGTGTTGGCGGGTGTTGGCGGGTGTTGGCGGGTGTTGGCGGGTGTTGGCGGGTGTTGGCGGGTGTTGGCGGGTGTTGGCGGGTGTTGGCGGGTGTTGGCGGGTGTTGGCGGGTGTTGGCGGGTGTTGGCGGGTGTTGGCGGGTGTTGGGGGG	න <u>ල</u> ට	Sau96 I Sau96 I Eco0109 I I Hae III I Hae III TAAGGCCTCGTGATACGCCTATTTTTATAGGTTAATGTCATGATAATAGGTTTC ATTCCCGGAGCACTATGCGGATAAAATATCCAATTACAGTACTATTATACAAAGTATTACAAAGTATTACAAAAGTATACAAAAAAAA	1942

1971		35,	/68			51
· ·	TTTTTCTAATACATTCAAATATGT AAAAAGATTTATGTAAGTTTATACA	Wbo II AGGAAGAGTATGAGTATTCAACATTTCCGT 2160 TCCTTCTCATACTCATAAGTTGTAAAGGCA	SfaN I A A G A T G C 2 2 4 0 T C T A C G 2 2 4 0	ogW	TCCTTGAGAGTTTTCGCCCC	2298 2298 Fig.
VI DIN 1943	II II AAATGTGGGGGGATGGGGATAAACAAT	2004 2004 2004 2005 2005 NIG III SSP I BSPH I LACCTGATAATGCTTCAATATTGAAAA	F NU 4H I F NU 4	ATTCCCTTTIIICCCCTAAACGGAAGGAACTAAAACGGAAGGAACAAAACGGCAAGGGAAAAACGCCGTAAAACGGAAGGAA	USE HgiA hspb 11 o I Bsp1286 I Mae III hill hill hspb 11 I ApdL I Hae III hill hill hold hill hill hill hill hill hill hill hi	2242 2245 2245 2245 2245

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2400 2480 2560 Fig.	4

•	37/68	,	15 2
2640	2720	2800	Fig.
Sau36 I Sau36 I Alw I Alw I Alw I Alw I Alw I Alw I Alu I	GAAGCCATACCAAACGACGAGCGTGACCACGATGCCT	MSP II Scrf I Scrf I Nci I NSE I Hha I Hha I Hha I Hor	2

	38/68		5M
FRU4H I NIG IV Bbv I NIG IV CTCGCGGTATCATTGCAGCGCAGGCGCATAGTAACGTCGTGACTAGTAACGTCGTGACTGAGTAACGTCGTGACTGAGTAACGTCGTGACTGAGTAACGTCGTGACTGAGTAACGTCGTGACTGAGTAACGTCGTGACTGAGTAACGTCGTGACTGAGTAACGTCGTGACTGAGTAACGTCGTGACTGAGTAACGTCGTGACTGAACGTCGTGACTGAACGTCGTGAACGTCGTGACTGAACGTCGTGAACGTGAACGTGAACGTGAACGTGAACGTGAACGTAACGTAACGTAACGTAACGTAACGTAACGTAACGTAACGTAACGTAACGTAACGTAACGTAACGTAACGTAACGTAACAACAACAACAACAACAAAAAAAA	FOK I AACTATGGATGAAGGAATAGACA 2960 TTGATACCTACTTGCTTTATCTGT 2943	MSe I Dra I 1 1 2 2 40 ACTCATATATACTITAGATIGATI 3040 TGAGTATATAGAATCTAACTAA 3039 3039 3040	Fig. E
	2843 2843 2845 Ple I Hinf I Hinf I ATCAATAGATGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	se I MOE III TAAGCATIGGTAACTGTCAGACCAAGITT ATTCGTAACÇATTGACAGTCTGGTTCAAA ATTCGTAACÇATTGACAGTCTGGTTCAAA 987 2997	Mae I Mbo I Dpn I
MSP I Sau96 I Hpd II CGCCCTTCCGCTGGTTTATTGC GCCGGGAAGGCCGACCGACCAAATAACG	28 28 20 20 20 20 20 20 20 20 20 20 20 20 20	U3A I MIG IV O I BON I WI TCGTGAGATAGGTGCCTCACTGAT AGCGACTCTATCCACGGAGTGACTA AGCGACTCTATCCACGGAGTGACTA 61 2974 2	2966

•	39/68	2 N
11 GAG 3120 CTC	Fnu4H I Bbv I TCTG 3200 AGAC AGAC 3200 3200	Mae III AAGG 3280 Fig.
MSe I MSe I CCCTTAACGT AGGGATTGCA 3111	Bstu I Hinp I Hha I CTGCGCGTAAI 3189 3189 3189	<u> </u>
SpH I I I I I I I I I I I I I I I I I I I	Squ3A I Mbo I Dpn I A W I S + Y I GATCCTTTTTT CTAGGAAAAA 174 3175 3175 3175	Mbo I Dpn I Alw I Sp II Pg II Alu I CGGATCAAGAGCTACCAACTCTTTTTCCGAAGG 3
TTTTGATAATCT	- 4543 M	MBO Dpn Alw I MSP I HPG II +h111 II
I BS 1 I BS 1 I I HPh I I I I I I I I I I I I I I I I I I I	Squ3A Mbo I Dpn I 17CTAGTT 3156 3156	NspB II CAGCGGTGGTTI
A I DPD A I BS FY A I BS FY A I I BS FY A I T T C C T A G A T T T T C C T T C T C T C T C T C T C	I TCAGACCCCGTAGAA AGTCTGGGGCATCTT	ACCACCGCTAC
MSB I D COLOR MSB I I I I I I I I I I I I I I I I I I I	Hga I Dde I TTCGTTCCACTGAGCGTC AAGCAAGGTGACTCGCAG AAGCAAGGTGACTCGCAG	THHIII II CTGCTTGCAACAAAAACCACCGCTACCAG
TAAAACTTO	TTTTCGTT AAAAGCAA	T+h111

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080	3360 I	1 3440	3520 Fio.	
·	HinP I Hha I CTGGCTTCAGCAGAGAGATACTGTCCTTCTAGTGAGTTAGGCCACCACTTCAAGAACTCT GACCGAAGTCGTCTCGCGTCTATGACAGATCACATCGGCATCAATCCGGTGGTGAAGTTCTTGAGA 3339	Fnu4H I hsp I hyp I hpv I hpv I hpp	NSPB II FRUM H I BBV I Hinp I Hinp I Hinp I Hinp I Hinp I Hing I CAACCTCAAGACGATAGCCCTATTCCGGTCGGCTTGCCCCCAAGCTCGGTCGAACCCCCCCC	

		42/68		50
	3840 0	3920	4000	Fig.
Hae III EcoR II Hae III NSPH I BS+N I Hae III NSP7524 I Aft III Aft III	ACGCCAGCAACGCGCCTTTTTACGGTTCCTGGCCTTTTGCTCACATGTTCTTTCCTGCGTTATCCCTTTGCTCCTTTCCTGCCTTTTTTTT	Fnu4H I Bbv I Bbv I I I I I I I I I I	Hinp I Ha I H	

	4	43/68	24
	I I I I 4080	4160	Fig. 5
3971 3976 3993	I MSe I ASE I ASE I ASE I ASE I ASE I SECONDERINATER ACCITANT ACACT CANTER A 4045 A 4046	S F V C C C C C C C C C C C C C C C C C C	
3942	HinP Hhg I HcGACAGGTTTCCCGACTGGAAAGCGGGCAGTGAGCGCA TGCTGTCCAAAGGGCTGACCTTTCGCCCGTCACTCGCGT Ho35	MSP II HPG III GCTTTACACTTTATGCTTCCGGCTCGTATGTTGTGTGG CGAAATGTGAATACGAAGGCCGAGCATACAACACACC 4099 4099 NIG III CCATGATTACGCC 4173	

44/68 HinD III Sph I 7 13 Pst I 15 BspM I 19 Sal I 25 Xba I 31 BamH I 36 Ava I 36 Sma I 36 Xma I 118 Asp718 118 Kpn I 299 Nco I ,326 581 Xca I Ser 65 → Thr 620 Hpa I 633 Ile167→Thr 850 Bsm I 859 EcoR I presumed gGFP-TT 4173 base pairs unique sites Fsp I 2702 Pvu I 2555 1674 Spe I Sca I 2444 1680 Eag I 1684 Spl I Ssp I 2120 1689 Apa I San II

1689

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Fig.

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Fig. 7A
Fig. 7B
Fig. 7C
Fig. 7D
Fig. 7E
Fig. 7F
Fig. 7G
Fig. 7H
Fig. 7I
Fig. 7J
Fig. 7K
Fig. 7L
Fig. 7M
Fig. 7N
Fig. 70
Fig. 7P
Fig. 7Q
Fig. 7R
Fig. 7S
Fig. 7T

> Fig. 7

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H	320	400		Fig
209 209 210 210 210	316 44 316 44 316	MDB I Xbd I Xbd I Xbd I Xbd I CALCETT AT A TACCETT CAAATAATACCCAACATTTGCTCTAATTGTGAAA CAATTTTGTGAAA CAATTTGTGAAA CAATTTGTGAAA CAATTTGTGAAA CAATTTGTGAAA CAATTTGTGAAA CAATTTGTGAAA CAATTTGTGAAA CAATTTGTGAAA CAATTTGTGAAAA CAATTTGTGAAAAAAAAAA	90	Ben I Ava I Sec I Sau3A I

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800	880 0	4 096	1040	Fig.
Squ3A I Nbo	Af I III Af I III GACGGGAACTACAAGACACGTGCTGAAGTTGAAG CTGCCCTTGATGTTCTGTGCACGTTCAAGTTTGAAG 880 856 856	GATGGAACATTCTTGGACACAAATTGGAA 96 CTACCTTTGTAAGAACCTGTGTTTAACCTT	AATGGAATCAAAGTTAACTTCAAAACTAGACA TACCTTAGTTTCAATTGAAGTTTGATTTGA	F

WU 90/209/1		•
	51/68	7.
1120	1200	1280 Fig.
CAACATTGAAGATGGAAGCGTTCAACTAGCAGACCATTATCAACAAATACTCCAATTGGCGATGGCCTGTCCTTTTAC GTTGTAACTTCTACCTTCGCAGTTGATCGTCTTTTATGAGGTTAACCGCTACCGGGACAGGAAATG GTTGTAACTTCTACCTTCGCAAGTTGATCGTCTTTTATGAGGTTAACCGCTACCGGGACAGGAAATG 11048	Sau3A I Mbo I Mbo I Bs i W I Avg II Avg II Avg II I N I G I I N I I I N I I I N I I I N I I I N I I I N I I I N I I I N I I I N I I I N I I N I I I N I I I N I I I N I I I N I I I N I I I N I I I N I I I N I I I N I I N I I I N I I N I I N I I N I I N I I N I I N I I N I I N I I N I I N I	FOK I Stop Stop I HinP

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1360	1440	1 1520 9 Fig.	0
NSPB II nu4H I nu4H I nu4H I nu4H I ge III ge I IV RS+U I RNI RS+U I NNI GGCGACATCTCGCGCCGTGCT GGCGACATCTCGCGCCGTGCT	1342 1320 1321 1322 1322 1323 HgiA I Bsp1286 I TCTCCCTGTGCTCCCCCTATTTTTGTTATCAAA AGAGGGACACGAGGGTGGGGATAAAAGTTTT	Hph I Mae III Xmn I Xmn I Hinf I Xmn II MSe I Mn! I Kml I MSE I Mn! I MSE I Kml I MSE I Kml I MSE I E Kml I E Kml I MSE I E Kml	

54/68	71	71
2000 2080 1111 2160	Fig.	111 2160 9 Fig.
AACCCCCAAAGGGGATAACAAGAGTTCTCAAAGGTCCTGCGGCAAAAGAAGAACGATTTTAGTGTTCATAACTA 1865 1880 1865 1882 1865 1882 1865 1882 1867 1867 1867 1874 1868 1960 1974 1974 1938 1943 1988 1964 1984 1964 1974 1974 1938 1943 1986 1964 1967 1974 1974 1974 1974 1974 1976 1974 1974 1974 1974 1974 1974 1974 1976 1974 1976 1974 1976 1974 1976 1974 1976 1974 1976 1974 1976 1977 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	TGCCGGGAAAGCAGAGCGAAAGCCACTACTGCAGACTGTGTACGTCGAGGGCGTCTGCA TGCCCGGGAAAGCAGAAGCCACTACTGCACTTTTGGAGACTGTGTACGTCGAGGGCGTCTGCAA	Saug6 I Saug6 I Saug6 I Saug6 I NI
CGAAAACCCCCAA P 1286 I S fan I 1 A I CGACGATGCAAGA CGTGCTACTTCT 21 21 1927 1927 1927 Sauge Sauge Sauge Sauge Sauge Sauge Sauge I I Hae I I B I Hae I I B I B B I I I B I B B I I I B I B B I I I B I I B B I I I I B B I I B I I	81 2090	Sau96

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	56/68			7K
2400	2480	2560 I	2640	Fig.
ATAATGGTTT TATTACCAAA	TTCAAATATG AAGTTTATAC	ACATT IGTAA	TAAAAGA ATTTCT	
BSPH I I I I I I I I I I I I I I I I I I I	TCTAAATACA AGATTTATGT	TATGAGTATTCA. ATACTCATAAGT HPh I	CGCTGGTGAAG GCGACCACTTTC/ 2624	
MSB HSB GGATAAAATATCCAATT 2377	IV CCCCTATTTGTTTATT GGGGATAAACAAATAA	P. 1 TATTGAAAAGGAAGAG ATAACTTTTTCCTTCTC 21 2534	TTTTGCTCACCCAGAAA AAAACGAGTGGGTCTTT 2609	
Squ96 I CO0109 I Hqe III CCCCGGAGCACTATGC CCCGGAGCACTATGC 2349 347 2351	III BS + U I I BS + U I I HinP I HinP I Hha I I I Hha I I I I Had I I I I I I I I I I I I I I I I I I I	GACAATAACC	GCA CGT 3	, !

	57/68	12	
2720	2800	2880 Fig.	€ .
Squ3A I HgiA I Bbh I Mbo I Mbo I Dpn	THE PLANT BE TO SERVING THE PROBLEM OF THE PROBLEM	RSa I Mae III Şfan I Fok I Sca I Mae III Şfan I Fok I Sca I Mae III Şfan I Fok I FOK I SCACTCGCCGCCGCCGCTATTCTCAGAATGACTTGGTTGAGTGCTTGCT	

	58/	⁷ 68		7.M
I 2960	3040	4	3120	Fig.
CCATAACCATGAGATGAGGCCGATTACTTCTGACAACGCTTGGTGATGGTGTGTGT	2882 2934 2935 2935 2935 2935 2935 2935 2935 2935	AACGTGTTGTACCCCTAGTACATTGAGCGGAACTAGCAACCCTTGGCCTC 2996	GACACCACGATGCCTGTAGCAATGGCAACGTTGCGCCAAACTATT CTGTGGTGCTACGGACATCGTTACCGTTGCAACGCGTTTGATAA . 3099 3103	

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59/68
                                                                                                         Fig.
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3273
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	61/68		7P
	3680	3760 I I I	3840 Fig.
3590 3590 3591	CCAACTCTTTTTCCGAAG GGTTGAGAAAAGGCTTC	GCCACCTTCAAGAACTC GCCACCTTCAAGAACTC CGGTGGTGAAGTTCTTGAG A0 741 Scr Msp Hpd	GATAAGTCGTGTCTTACCG CTATTCAGCACAGAATGGC
-3575 3576 3576 3576 3576	00 I 01 I 1 Alu I ATCAGAGCTA TAGTTCTCGAT 3658 3658	151 Hq Hq 100 100 11 11 11	CTGCCAGTGGC GACGGTCACCG
3564 3564 3564 3567 35655 35655	Mbe DPI	Mae I TTCTAGTGTA AAGATCACAT 3723 3723 Fnu	CAGTGGCTG GTCACCGAC
3557 3557 3557	NspB II CCAGCGGTGGT GGTCGCCACCA	AAATACTGTO	CTAATCCTGTT GATTAGGACAA
3533	Tthiii II Bby I Tthii II GCTGCTTGCAAACAAAAACCACCGCTAC GCTGCTTGCAAACAAAAAAAACCACCGCTAC GCACGAACGTTTTTTTGGTGGCGATG CGACGAACGTTTTTTTGGTGGCGATG 3601 3609	CATACC CTATACC	MnI I TGTAGCACCGCCTACATACCTCGCTCTGC ACATCGTGGCGGATGTATGGAGGCGAGACC

	62	2/68		70
 	3920	4000	4080	Fig.
3797 3802 3810 3807 3807 3810 3838 3838 3838 3810 3838 3838 3838	NSP I Hinf I Holl I Hinp I Alu I Apal I Alu I Apal I Hha I Apal I Alu I Apal I Hha I Apal I Alu I Apal I Alu I Apal I Alu I Apal I Alu I Apal I Apal I Alu I Apal I Apal I Apal I Alu I Apal I Apal I Apal I Alu I Apal I A	HinP Hha I Hae II CGTGAGCATTGAGAAAGCGCC GCACTCGTAACTCTTTCGCGC GCACTCGTAACTCTTTCGCGG 3973	80 00 0	

WO 98/28971				
		63/68		7R
1	4160	4240	I 4320	Fig.
4046 4056 4056 4056 4056 4056 4056	H90 I		Hinf I Hinf I Hinf I Alu I Fnu4H I Bbv I Hinf I Hinf I FGATTCTGTGGATAACCGTATTACCGCTTTGAGCTGATACCGCTCGCCGCAGCGCAGCGAGCG	

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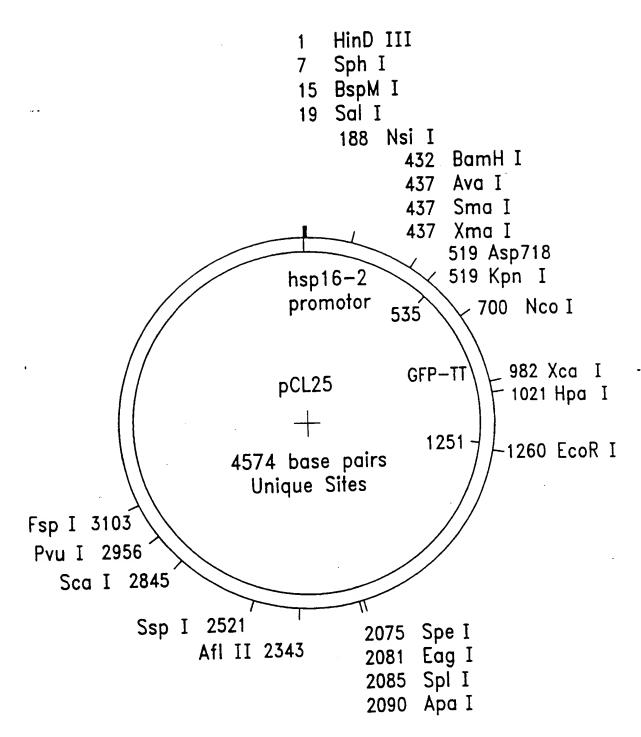


Fig. 8

aggaccagat gggcacgatg gacaagacgg agtcccagga gaccaccagg agaggctggg gccaggagac aggaaaggga ggatgaaaaa aatggagctc caggactcag aggaatgcgc ggagatgctg gatgettate caggacagte tggaccaeda accaaatgga accaggacca gtccattccc gaatggagga tggtgttcag ccaccagcac caactccaaa cccatatgga aagatgeega agatgtteaa aaeaeeeeae eaaeaggatg etteaeetgt aaatataaaa atttcagacc agaaggacca ccaggagccc iggitaaact iggagcagga accgeticca accgigigag acgicaacad egeaaattee aattgtagae aaatteatga eattttteea tgagctggat gccgaaatcg ccaacttcag agtgagtttc agatticcag atgaccetaa etaeggegae gieeggegee tttetaatte actgataatt teaattattt agagteteae tgaagaeatg aaaattitti gtittgtatt tggtttacca tggatgttaa ctttatcatt tcactttttc ttggcactcg tccaaaaatc tcaattaaaa atttctggat acccaggaat gccgaatact gcaaatgccc aggaagagaa atatggagga gctgctggag aggacgggta Highigati tetetititig ecgetgette getttaeagt gatctccagg atcacaatga catcettatg tatatgcatt ctttttattg tttcctgata tttattctct cagatggtat actcataccc agagacggaa ggaccaagag ggaaagccag gagaaattcc actgttgaga aggaggtgga ttcccagatg attaaaatac aaaactgtat aactggaaga acaaggacca gggccgcccd acgtccagga agctccacca atatcaatgc aggatccgat aaggagagga cacagggacc gccttcaagg gaggaaatca aggaaaagat caagccagcc gacaaccagg ეეეენნნნნე agagaccatt cacttggacc cagacgicat caaagtteee atggagccac catacticto ggggaaaata atatctggaa gacttettgt ctggagccac atctaactga ggagagccag ccagaaggac aaaaataaaa gacgaagtgc ccacaaggac gacgatggag ggaaatcaac aatattaaac caggacttcc gctggaccag gtggaccac ggaccacgtg ggagctcgtg ggatatggag Hgacggaa ctatatagca atggaggat caagtttcca gaactttett atatetttag lgggttgata attgtatttt .96 48 541 90, 301 24, $\dot{\infty}$ ؽ 7

Fig. o

DCGPPGAPGSDGKPGSPGGKGDDGERPLGRPGPRGPPGEAGPEGPQGPTGRDAYPGQSG KFPGGGFPDGPFPNGGGPRGGNQCQCTVENSCPPGPAGPEGEEGPDGHDGQDGVPGFD GKDAEDVQNTPPTGCFTCPQGPLGPQGPNGAPGLRGMRGARGQPGRPGRDGNPGMPG **LGAGTASNRVRRQQYGGYGATGVQPPAPTPNPYGGYGASQPAPPEKFPDGIPNGGNQP** PQGEPGLQGYGGAGEDGPEGPPGAPGLPGKDAEYCKCPGREGDAGRSARRHRKFQL MTLTTATSGAIVFSGATLLVSLFAAASLYSQVSNIWNELDAEIANFRSLTEDMWVDMVK

 ${
m Fig.}$ 10

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With international search report.

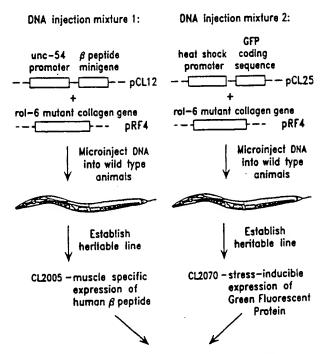
Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.

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(54) Title: TRANSGENIC ANIMALS FOR IN VIVO VISUALIZATION OF BETA-AMYLOID TOXICITY

(57) Abstract

The present invention relates to methods and compositions for visualization of the toxic effects of transgenes in vivo. In particular, the present invention provides methods and compositions for the production and use of transgenic, including dually transgenic, Caenorhabditis elegans for visualization of the toxic effects of \(\beta\)-amyloid accumulation in vivo.



Mate lines together, recover dual transgenic animals by classic genetic techniques

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A. CLASSIF	FICATION OF SUBJECT MATTER C12N15/00 A01K67/033 C07K14/43	35 C07K14/47	
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According to	International Patent Classification(IPC) or to both national classification	on and IPC	
	SEARCHED		
IPC 6	cumentation searched (classification system followed by classification AOTK CO7K		
Documental	tion searched other than minimum documentation to the extent that suc	ch documents are included in the lields sea	rched
Electronic d	ata base consulted during the international search (name of data base	e and, where practical, search terms used)	
C. DOCUM	ENTS CONSIDERED TO BE RELEVANT		
Category	Citation of document, with indication, where appropriate, of the rele-	vani passages	Relevant to claim No.
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X	cited in the application see the whole document	-/	11-15
X Fur	ther documents are listed in the continuation of box C.	X Patent family members are listed	in annex
	ther documents are listed in the continuation of box C. alegories of cited documents	"T" later document published after the inte	rnational filing date
consi E" earlier	nent defining the general state of the art which is not idered to be of particular relevance document but published on or after the international	or priority date and not in conflict with cited to understand the principle or the invention. "X" document of particular relevance, the	ine application but teory underlying the claimed invention
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later	P document published prior to the international filing date but after than the priority date claimed "3" document member of the same patent family		
	3 September 1998	Date of mailing of the international se	
	making address of the ISA European Patent Office, P.B. 5818 Patentiaan 2	Authorized officer	
	NL - 2280 HV Rijswyk Tel (+31-70) 340-2040. Tx 31 651 epo nl. Fax: (+31-70) 340-3016	Chambonnet, F	

Int :tional Application No PCT/US 97/23819

C.(Continu	ation) DOCUMENTS CONSIDERED TO BE RELEVANT	Relevant to claim No
Catedoo	Citation of document, with indication where appropriate of the relevant passages	
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Int. Ational application No. PCT/US 97/23819

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)
This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
1. Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:
~•
2. Claims Nos.: because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful international Search can be carried out, specifically:
3. Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6 4(a).
Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)
This International Searching Authority found multiple inventions in this international application, as follows:
see additional sheet
1. As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2. As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
Remark on Protest The additional search fees were accompanied by the applicant's protest. X No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

This International Searching Authority found multiple (groups of) inventions in this international application, as follows:

and partially 11-23 1. Claims: 1-10

a transparent animal comprising:

(a) a toxic transgene:

- (b) an inducible promotor operably linked to a reporter gene; wherein the toxic transgene induces the promotor and wherein the expression of the reporter gene is detectable in vivo; and a method for providing such a dually transgenic transparent non-human animal comprising:
 - a) providing:
 - i) a first and a second non-human transparent animals;
 - ii) a first toxic transgene; and
 - iii) a second transgene, comprising an inducible promotor operably linked to a reporter gene, wherein the toxic transgene induces the promotor and wherein the expression of the reporter gene is detectable in vivo,
 - b) introducing said first toxic transgene into the genome of said first non-human animal to produce a first transgenic animal and introducing said second transgene into the genome of said second non-human animal to produce a second transgenic animal; and
 - c) mating said first transgenic animal with said second transgenic animal to produce a dually transgenic transparent non-human animal wherein said toxic transgene is expressed and induces the promotor so that the said reporter gene is also expressed and detectable in vivo;
 - 2. Claims: partially 11-23

a method to produce a dually transgenic non-human animal with a toxic transgene and a reporter are expressed comprising:

- a) providing:
- i) a first and a second non-human animal;
- ii) a first toxic transgene; and
- iii) a second transgene, comprising a reporter,
- b) introducing said first toxic transgene into the genome of said first non-human animal to produce a first transgenic animal and introducing said second transgene into the genome of said second non-human animal to produce a second transgenic animal; and

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

c) mating said first transgenic animal with said second transgenic animal to produce a dually transgenic non-human animal wherein said toxic transgene and said reporter are expressed

in the cases non relevant of the first described subject, that means where it is not a transparent animal comprising :

(a) a toxic transgene;

(b) an inducible promotor operably linked to a reporter gene; wherein the toxic transgene induces the promotor and wherein the expression of the reporter gene is detectable in vivo.

Information on patent family members

Ir. ational Application No
PCT/US 97/23819

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